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SYNTHESIS AND BIOLOGICAL ACTIVITY OF NEW SUGAR MODIFIED 2',5'-OLIGOADENYLATE TRIMERS

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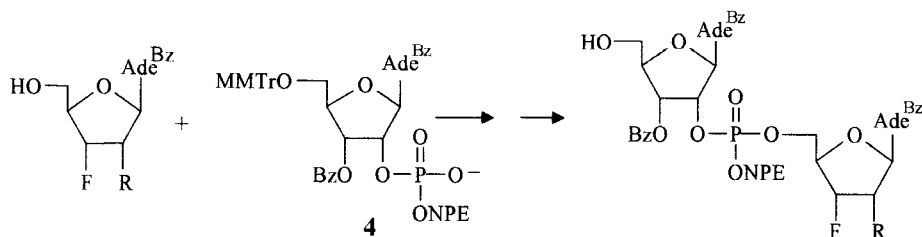
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ABSTRACT: New 2',5'-oligonucleotide trimers containing 2',3'-dideoxy-3'-fluoro derivatives of adenosine at the 2'-terminal end of oligomers have been synthesized. Some of the synthesized trimers showed biological inhibitions of HIV-1 reverse transcriptase and HIV-1 induced syncytia formation.

Naturally occurring 2',5'-oligoadenylates (mainly trimers, both 5'-phosphorylated and unphosphorylated) have shown different kinds of biological activity¹⁻⁴. However, these compounds are rapidly inactivated in the cell by some enzymes^{5,6}. Many analogues of 2',5'-oligoadenylates have been synthesized to overcome this problem, and to achieve new approaches to antiviral and antitumoral therapy⁷⁻¹⁵. One of this analogues, cordycepin trimer core^{16,17}, was found to be a biologically active compound with metabolic stability¹⁸. Later it was found to be also an inhibitor of HIV-1 reverse transcriptase (RT)^{19,20}. As far as each individual nucleoside residue of 2',5'-oligoadenylates may assume a different role in inhibition of RT or RNase L activation, we have synthesized some new 3'-fluoro containing trimers with double modification at the 2',3'-terminus of the oligos, as potential metabolic stable 2',5'-oligoadenylates. The ability of synthesized trimers to inhibit HIV-1 replication and to improve RNase L activation have been also investigated.

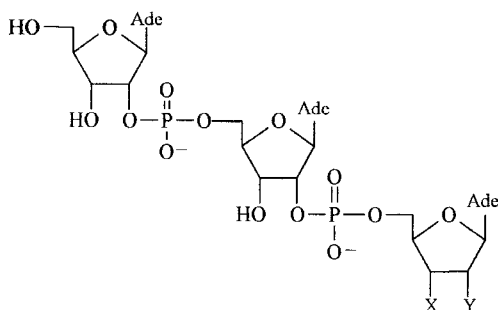
Selectively blocked nucleosides 1-3, used for the synthesis of trimers, have been obtained from the corresponding unprotected nucleosides²¹⁻²³, respectively, using the transient protection method²⁴. The synthesis of the 2'-phosphodiester 4 has been described by us earlier²⁵. Condensation of the 2'-terminal units 1-3 and the building block 4 in the presence of TPSCl/tetrazole 1:3, as catalysts and followed by detritylation with 2% TsOH in CH₂Cl₂/MeOH 7:3 afforded the 5'-OH containing dimers 5-7, respectively. The final condensation of 5-7 and 4, followed by treatment with 0.5 M DBU/Py, TsOH/CH₂Cl₂/MeOH and conc. NH₄OH led to the

corresponding trimers **8-11** which were isolated by ion exchange chromatography with overall yields of 40-50% from all steps. Catalytic hydrogenolysis of the azido derivative **8** in the presence of 10% Pd/C in EtOH gave 2'-amino-3'-fluoro-2',3'-dideoxy containing trimer **11**.



N	R
1	N ₃
2	H
3	Cl

N	R
5	N ₃
6	H
7	Cl



N	X	Y
8	F	N ₃
9	F	H
10	F	Cl
11	F	NH ₂
12	OH	OH

The synthesized trimers were tested for recombinant human GST-RNase L activation, and inhibition of RT and HIV-1 induced syncytia formation by assays described previously²⁶. Summarized data in comparison to the activity of the 2',5'-oligoadenylate trimer core **12**, are presented in the following table.

Compound **9-11** inhibited induced syncytia formation 7.0, 2.0, and 9.0 fold, respectively, compared to 3.0 fold reduction with unmodified trimer **12**. Trimers **8-10** inhibited HIV-1 RT activity 99.7%, which compares with a 33% inhibition of HIV-1 RT by the compound **12**. The previously obtained data about a 96% inhibition of HIV-1 RT by the cordycepin trimer core²⁷ and the results presented here, show that hydroxyl group at either the C2' or C3' position of the trimer **12** are not essential for the inhibition of HIV-1 RT activity. When 2',5'-oligoadenylate **12** is modified at the C2' or C3' position either fluorine, chlorine, azido and amino substituent, recombinant human GST-RNase L is not activated to hydrolyze poly(U)-3'-[³²P]pCp compared to a 50% activation of GST-RNase L by the trimer **12**. These results, and the data a 12% activation

TABLE. Biological activities of the synthesized trimers.

Comp.	X	Y	RNase L ^a	Syn ^b	RT ^c
8	F	N ₃	0	-	99.7
9	F	H	0	7.0	99.7
10	F	Cl	0	2.0	99.7
11	F	NH ₂	0	9.0	-
12	OH	OH	50	3.0	33.0

^a- RNase L is the percent hydrolysis of [³²P]poly(U) in the presence of 10 nM p₃A₃ and 10 μM for the remaining trimers.
^b- Syn is the fold reduction in HIV-1 replication, as determined by Syncytia formation.
^c-RT is the percent inhibition of HIV-1 RT.
The value given in the mean of duplicate determination with a variance of ±5-10%.

of GST-RNase L by the both cordycepin trimer core and its conjugate with vitamin E at the 2'-terminus of the trimer²⁷, indicate that the oxygen of the hydroxyl group at the C2'-position of the 2',3'-terminus is essential for the activation of GST-RNase L.

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REFERENCES

1. Kimchi A., Shure H., Revel M. *Nature* (London). **1979**, 282, 849.
2. Leanderson T., Nordfelth R., Lundgren E. *Biochem. Biophys. Res. Commun.* **1982**, 107, 511.
3. Devash Y., Biggs S., Sela J. *Science* **1982**, 216, 1415.
4. Devash Y., Gera A., Willis D.H., Reichman M., Pfeleiderer W., Charubala R., Sela I., Suhadolnik R.J. *J. Biol. Chem.* **1984**, 259, 3482.
5. Eppstein D.A., Barnet J.W., Marsh Y.Y., Gosselin G., Imbach J.L. *Nature* (London). **1987**, 302, 723.
6. Herdewijn P., Charubala R., De Clercq E., Pfeleiderer W. *Helv. Chim. Acta.* **1989**, 72, 1739.
7. Kwiatkowski M., Gioelli C., Chattopadhyaya J.B., Oberg B., Drake A.F., *Chem. Scripta* **1982**, 19, 49.
8. Mikhailov S.N., Pfeleiderer W. *Tetrahedron Lett.* **1985**, 26, 2059.
9. Herdewijn P., Charubala R., Pauwels R., De Clercq E., Pfeleiderer W. *Nucleosides & Nucleotides* **1987**, 6, 443.
10. Kariko K., Sobol R.W., Suhadolnik L., Li S.W., Reichenbach N.L., Suhadolnik R.J., Pfeleiderer W. *Biochemistry* **1987**, 26, 7127.
11. Herdewijn P., Charubala R., Pfeleiderer W. *Helv. Chim. Acta.* **1989**, 72, 1729.
12. Charubala R., Pfeleiderer W., Sobol R.W., Li S.W., Suhadolnik R.J. *Helv. Chim. Acta.* **1989**, 72, 1354.

13. Kitade Y., Nakata Y., Hirota K., Maki Y., Pabuccuoglu A., Torrence P.F. *Nucl. Acids Res.* **1991**, *19*, 4103.
14. Pfeleiderer W., Himmelsbach F., Charubala R. *Bioorgan. Med. Chem. Lett.* **1994**, *8*, 1047.
15. Kvasyuk E.I., Kulak T.I., Mikhailopulo I.A., Charubala R., Pfeleiderer W. *Helv. Chim. Acta.* **1995**, *78*, 1777.
16. Charubala R., Pfeleiderer W. *Tetrahedron Lett.* **1980**, *21*, 4077.
17. Charubala R., Uhlmann E., Himmelsbach F., Pfeleiderer W. *Helv. Chim. Acta.* **1987**, *70*, 2028.
18. Doetsch P.W., Suhadolnik R.J., Sawada Y., Mosca J.D., Flick M.B., Reichenbach N.L., Dang A.Q., Wu J.M., Charubala R., Pfeleiderer W., Henderson E.E. *Proc. Natl. Acad. Sci. U.S.A.* **1981**, *78*, 6699.
19. Müller W.E.G., Weiler B.E., Charubala R., Pfeleiderer W., Leserman L., Sobol R.W., Suhadolnik R.J., Schröder H.C. *Biochemistry*, **1991**, *30*, 2027.
20. Schröder H.C., Suhadolnik R.J., Pfeleiderer W., Charubala R., Müller W.E.G. *Int. J. Biochem.* **1992**, *24*, 55.
21. Mikhailopulo I.A., Poopeiko N.E., Pricota T.I., Sivets G.G., Kvasyuk E.I., Balzarini J., De Clercq E., *J. Med.Chem.* **1991**, *34*, 2195.
22. Chiedgeavadze Z.G., Beabealashvilli R.S., Scamrov A.V., Kvasyuk E.I., Zaitseva G.V., Mikhailopulo I.A., Kowollik G., Langen P. *FEBS Lett.* **1985**, *183*, 275.
23. 2',3'-Dideoxy-2'-chloro-3'-fluoroadenosine have been obtained as a gift from Dr. Tamara Pricota (Institute of Bioorganic Chemistry Byelorussian Academy of Sciences, Minsk).
24. Ti G.S., Gaffney B.L., Jones A.A. *J. Am. Chem. Soc.* **1982**, *104*, 1316.
25. Kvasyuk E.I., Kulak T.I., Khripach N.B., Mikhailopulo I.A., Uhlmann E., Charubala R., Pfeleiderer W. *Synthesis* **1987**, 535.
26. Wasner M., Suhadolnik R.J., Horvath S.E., Adelson M.E., Kon N., Guang M.-X., Henderson E.E., Pfeleiderer W. *Helv. Chim. Acta.* **1996**, *79*, 609.
27. Wasner M., Suhadolnik R.J., Horvath S.E., Adelson M.E., Kon N., Guang M.-X., Henderson E.E., Pfeleiderer W. *Helv. Chim. Acta.* **1996**, *79*, 619.